



## Resin-bound mercapto acids: synthesis and application

Spyros Mourtas, Dimitrios Gatos, Manolis Karavoltzos, Christina Katakalous and Kleomenis Barlos\*

Department of Chemistry, University of Patras, Patras, Greece

Received 26 February 2002; accepted 7 March 2002

**Abstract**—Mercapto acids were attached through their thiol group onto 2-chlorotrityl (Clt)-, trityl (Trt)-, 4-methyltrityl (Mtt)-, 4-methoxytrityl (Mmt)- and 4,4'-dimethoxytrityl (Dmt)-resins. The new resins were used in the solid-phase synthesis of small mercaptoacylamino alcohols. Cleavage from the resins was performed by treatment with trifluoroacetic acid (TFA) solutions in dichloromethane (DCM) using triethylsilane (TES) as scavenger. © 2002 Elsevier Science Ltd. All rights reserved.

Mercaptoacyl aminoacids are inhibitors of several metallopeptidases, such as the antihypertensive captopril **1**<sup>1,2</sup> and the analgesic thiorphane **2**<sup>3,4</sup> (Fig. 1). In addition, mercaptoacyl dipeptides are strong dual peptidase inhibitors.<sup>5–7</sup> In order to synthesize new lead structures using solid-phase combinatorial methods, suitably resin-bound mercapto acids are required.

The Trt- and Mmt-groups, are useful protecting groups for the thiol-group of cysteine.<sup>8</sup> Therefore, we applied resins of the trityl-type **3**,<sup>9–11</sup> for the attachment of mercapto acids<sup>12</sup> through their thiol group. Thus, a two-fold molar excess of **4** and less than an equimolar amount of diisopropylethylamine (DIPEA) was left to react with resins **3** for 0.5–2 h at rt in DCM/dimethylformamide (DMF) (1:1) (Scheme 1). Under these conditions the attachment of **4** through their carboxy group is not possible. To ensure that thiols **4** were exclusively bound to the various resins by their thiol function, **5** were treated with a mixture of acetic acid (AcOH)/TFE/DCM (1:2:7) for 15 min at rt. Under these conditions the resin-bound carboxy group is cleaved quantitatively, even in the case of the most acid stable resin **5e**. Unreacted remaining trityl chlorides were converted to the corresponding inert trityl-methyl ethers, by washing

resins **5** with methanol (MeOH)/DIPEA/DCM (15:5:80). The loading of the resins obtained was determined by sulfur analysis. We observed highest attachment rates in the case of **5a–b**. Thus, resins **5a–b** were obtained within 30 min with a loading of 0.7–0.8 mmol **4**/g. In contrast, **5e** gave at the same time resins with a loading of 0.4–0.6 mmol **4**/g. Besides resin-bound linear mercapto acids **5**, we obtained in excellent yield the sterically hindered thiolactic acid **6** and the *m*- and *p*-mercaptomethyl benzoic acids **7**, **8**.

The cleavage of the mercapto acids from the various resins was performed by treatment with TFA solutions in DCM/TES (97:3). In general the acid sensitivity of the mercapto-resin bond seems to be independent of the bound mercapto acids. The acid sensitivity of the resin-bound mercapto acids increases considerably from **5e** to **5a**. Thus, complete cleavage of the mercapto acids from the resins **5a–e** was effected by 4×3 min treatment

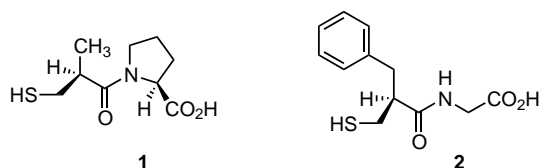
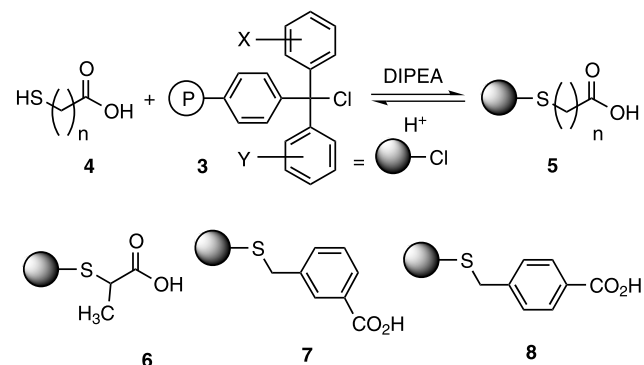


Figure 1.



**3**; X, Y = 4-CH<sub>3</sub>O, 4-CH<sub>3</sub>O (**a**), 4-CH<sub>3</sub>O, H (**b**), 4-CH<sub>3</sub>, H (**c**), H, H (**d**), 2-Cl, H (**e**), n = 1–5

\* Corresponding author.

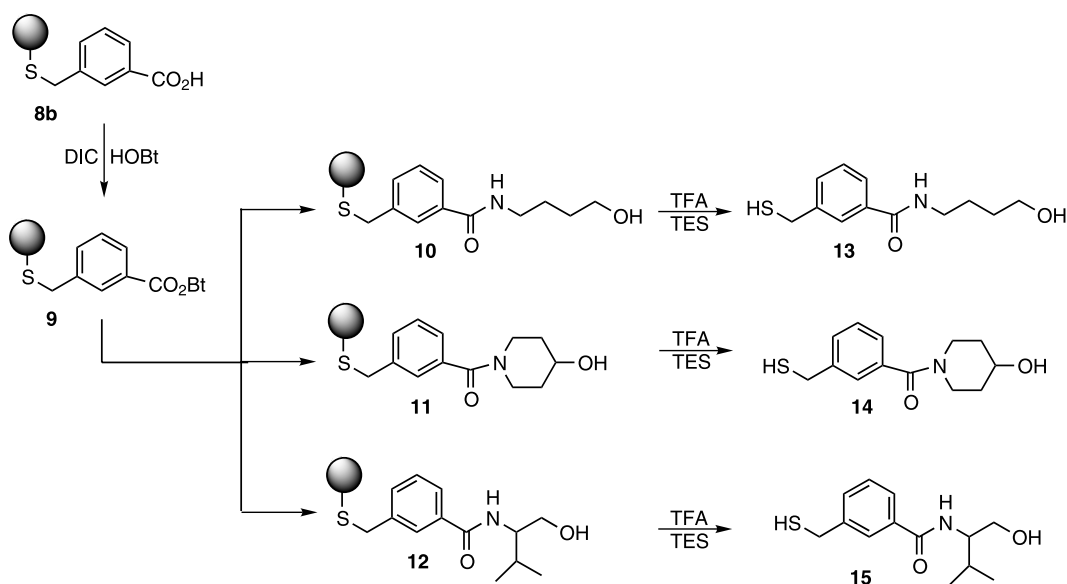
Scheme 1.

with 0.5, 1.1, 10, 30 and 65% TFA, respectively in DCM/TES (97:3).

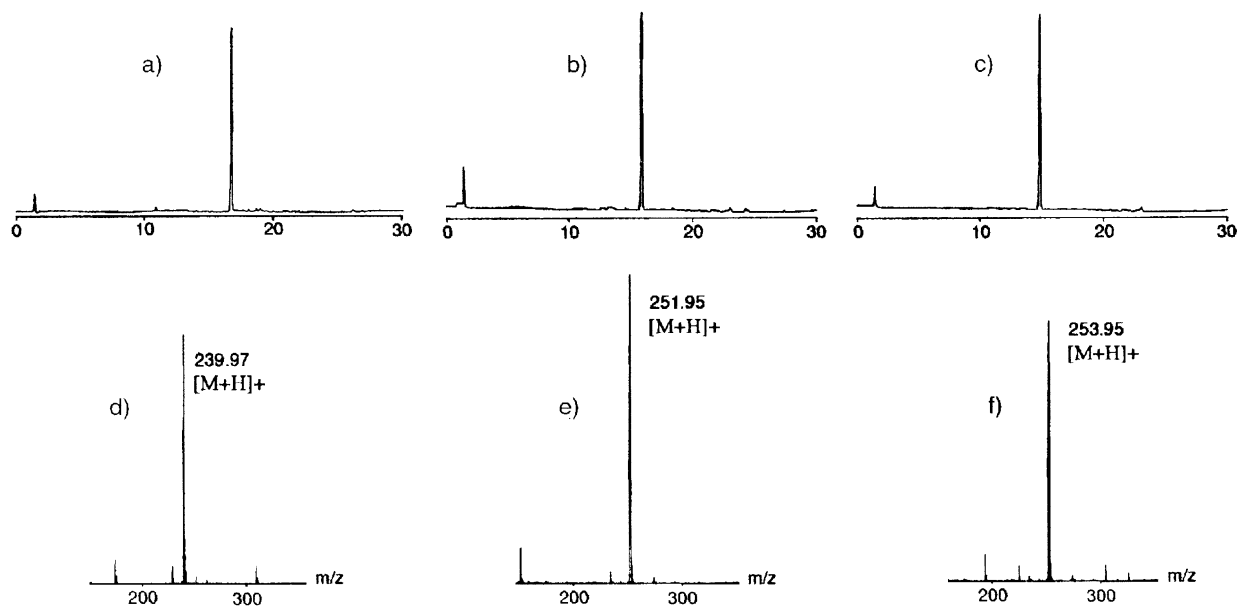
The utility of resin-bound mercapto acids in solid-phase synthesis was investigated during their coupling with amino components. Thus, **5–8** were converted to the corresponding benzotriazolyl esters, by treatment with excess diisopropylcarbodiimide (DIC)/1-hydroxybenzotriazole (HOBt) in tetrahydrofuran (THF) for 4 h at rt. The resin-bound mercapto acid benzotriazolyl esters obtained were reacted with a three-fold molar excess of amino acid esters and aminoalcohols (linear, cyclic and derived from amino acids). In one example, **8b** was converted to the active ester **9** and reacted subsequently

in DMF, for 1 h at rt with aminobutanol, 4-hydroxypiperidine and valinol, to yield the resin-bound mercaptoacylamino alcohols **10–12** (Scheme 2). Treatment with 1.1% TFA in DCM/TES (97:3) for 4×3 min, released the crude alcohols **13–15** in 89, 94 and 92% yields and 94, 92 and 94% purity, respectively, according to HPLC analysis (Fig. 2a–c). Their correct molecular weight was determined by ES–MS (Fig. 2d–f).

The conversion of the resin-bound mercapto acids to the corresponding benzotriazolyl esters and mercaptoacylamino alcohols were directly monitored by FT-IR analysis. As an example, the conversion of **8b** to the benzotriazolyl ester **9** was observed by the complete



Scheme 2.



**Figure 2.** Analytical HPLC of crude **13** (a), **14** (b) and **15** (c); column: Lichrospher RP-8, 5  $\mu$ m; 4×150 mm; gradient: from 20 to 100% B acetonitrile in water within 30 min; flow rate 1 ml/min; detection at 254 nm; ES–MS at 30 eV of purified **13** (d), **14** (e) and **15** (f).

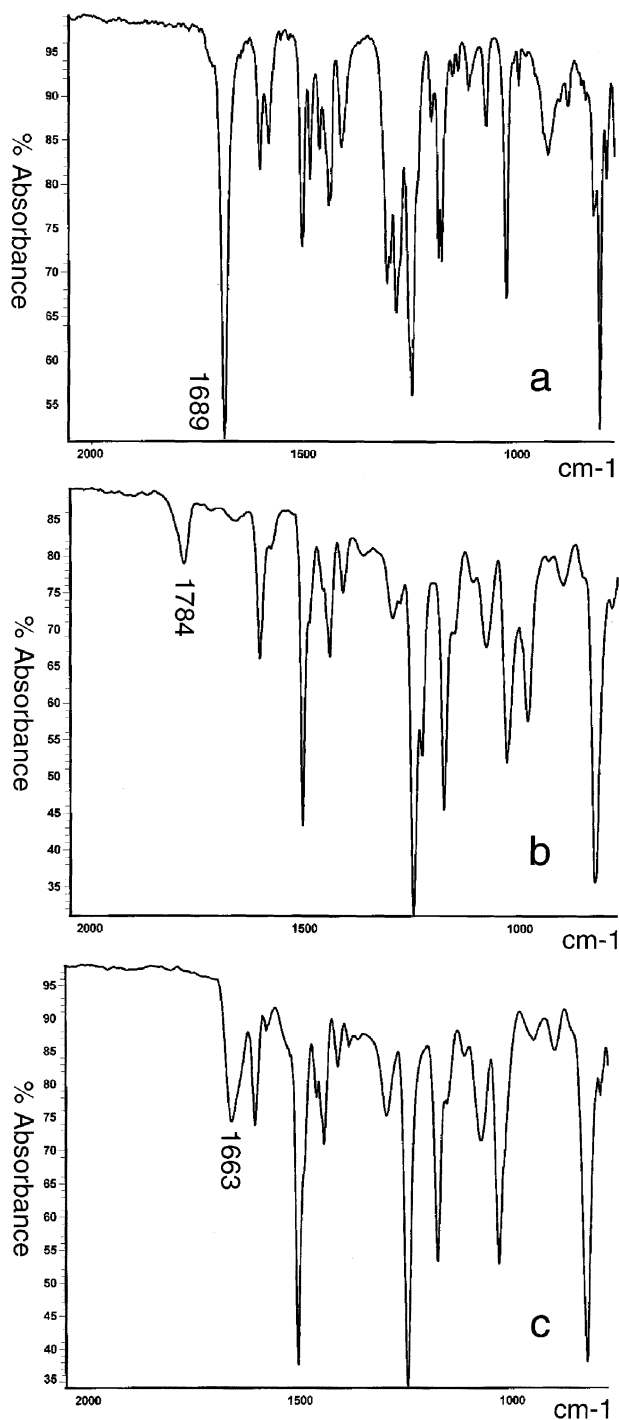


Figure 3. FT-IR analysis of **8b** (a), **9** (b) and **12** (c).

disappearance of the carboxy carbonyl frequency at  $1689\text{ cm}^{-1}$  and the appearance of an ester carbonyl frequency at  $1784\text{ cm}^{-1}$  (Fig. 3a–b). The second reaction is the nucleophilic replacement of the hydroxybenzotriazolyl group of **9** by the amino group of valinol. The success of this step is shown by the appearance of the amide carbonyl band at  $1663\text{ cm}^{-1}$ . The conversion was complete as seen by the disappearance of the ester carbonyl band at  $1784\text{ cm}^{-1}$  (Fig. 3b–c).

#### Acknowledgements

The authors acknowledge CBL-Patras S.A. for financial support.

#### References

- Ondetti, M. A.; Rubin, B.; Cushman, D. W. *Science* **1977**, *196*, 441–444.
- Cushman, D. W.; Cheung, H. S.; Sabo, E. F.; Ondetti, M. A. *Biochemistry* **1977**, *16*, 5484–5491.
- Roques, B. P.; Fournié-Zaluski, M. C.; Soroca, E.; Lecomte, J. M.; Malfroy, B.; Llorens, C.; Schwartz, J. C. *Nature* **1980**, *288*, 286–288.
- Llorens, C.; Gacel, G.; Swerts, J. P.; Perdrisot, R.; Fournié-Zaluski, M. C.; Schwartz, J. C.; Roques, B. P. *Biochem. Biophys. Res. Commun.* **1980**, *96*, 1710–1716.
- Fink, C. A.; Qiao, Y.; Berry, C. J.; Sakane, Y.; Ghai, R. D.; Trapani, A. J. *J. Med. Chem.* **1995**, *38*, 5023–5030.
- Robl, J. A.; Karanewsky, D. S.; Asaad, M. M. *Tetrahedron Lett.* **1995**, *36*, 1593–1596.
- De Lombaert, S.; Blanchard, L.; Stamford, L. B.; Sakane, Y.; Berry, C.; Ghai, R. D.; Trapani, A. J. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 2875–2880.
- Barlos, K.; Gatos, D.; Hatzi, O.; Koch, N.; Koutsogianni, S. *Int. J. Pept. Protein Res.* **1996**, *47*, 148–153.
- Fréchet, J. M. J.; Maque, K. E. *Tetrahedron Lett.* **1975**, 3055–3058.
- Barlos, K.; Gatos, D.; Kallitsis, J.; Papaphotiu, G.; Sotiriu, P.; Wenqing, Y.; Schäfer, W. *Tetrahedron Lett.* **1989**, *30*, 3943–3946.
- Barlos, K.; Gatos, D.; Chatzi, O. In *Peptides; Proceedings of the 22nd European Peptide Symposium 1992*; Schneider, C. H.; Eberle, A. N., Eds.; ESCOM: Leiden, 1993; pp. 281–282.
- Mourtas, S.; Gatos, D.; Kalaitzi, V.; Katakalous, C.; Barlos, K. *Tetrahedron Lett.* **2001**, *42*, 6965–6967.